

**Modular On-Demand Water Purification for Developing Countries**

2021-2022 Challenge Water Testing Procedures (updated 11.03.22)

**Challenge Water Makeup**

The challenge water will have the characteristics/components listed in table 1, shown below.

Table 1: Challenge Water Characteristics/Components

|  |  |  |  |
| --- | --- | --- | --- |
| **Contaminant** | **Reduction/ Maximum Level** | **Notes** | **Contaminant** |
| Bacteria (Brewer’s yeast surrogate) | Log reduction of 6 | 99.9999% removal12 | Bacteria (Brewer’s yeast surrogate) |
| Virus (no surrogate) | Log reduction of 4 | 99.99% deactivation12 | Virus (no surrogate) |
| Oocyst (Polymer microspheres surrogate) | Log reduction of 3 | 99.9% removal12 | Oocyst (Polymer microspheres surrogate) |
| Chlorine (if applicable) | 4.0 mg/L;  0.8 mg/L;  250 mg/L | As Cl2 or chloriamines; as chlorine dioxide6; as chloride9 (see problem statement if other disinfectant is used) | Chlorine (if applicable) |
| pH | 7.0 ± 0.5 | Secondary water regulation9 | pH |
| Turbidity | 5 NTU | Recommendation from the Sphere handbook11 | Turbidity |

**Required Equipment & Materials (with images)**

|  |  |
| --- | --- |
| Micropipettes (0.5 μL- 10 μL) and pipette tips | C:\Users\rubit\Desktop\eppendorf_research_10ul.jpg2-200 µl Micropipette Tips • MiniOne Systems |
| Beakers | C:\Users\rubit\Desktop\61Yo4dNgb3L._SL1500_.jpg |
| Graduate Cylinders |  |
| Light Microscope | C:\Users\rubit\Desktop\Light_micro.jpg |
| Hemocytometer | C:\Users\rubit\Desktop\Microscope-hemocytometer.jpg |
| Eppendorf tube | C:\Users\rubit\Desktop\download i.jfif |
| Hotplate & Magnetic Stirrer | C:\Users\rubit\Desktop\61BCDRhx3YL._SL1000_.jpg |
| Turbidity meter kit |  |
| Digital Balance | C:\Users\rubit\Desktop\B1155252604.jpg |
| Luer-Lok Filter | Cole-Parmer CA Syringe Filters, 0.20 um, 26 mm Dia; 100/Box |
| Sterile Syringe | https://assets.fishersci.com/TFS-Assets/CCG/Fisher-Scientific/product-images/Syringe_14955459_P1_Silo.tif-650.jpg |
| Chlorimeter |  |

**Addressing Criteria**

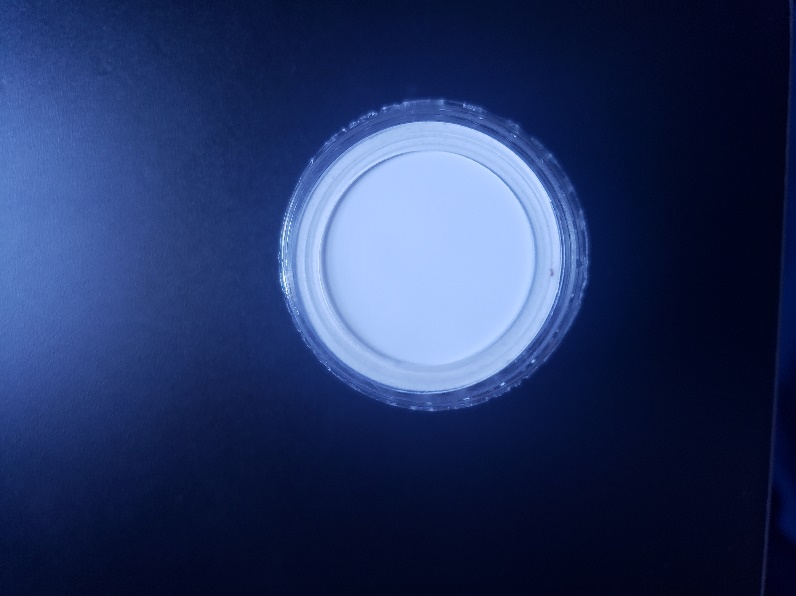
After a duel has been completed, ChemE Cube staff will collect the product water from both teams. The water will be tested to determine if criteria on the rubric have been met. The criteria and how they will be determined are the following:

*Criterion #1: “Product meets or exceeds 6 log reduction of bacterial surrogate”*

**Category:** Product

**Method:** Cell count though hemocytometer or filter.

**Procedure:**

1. Ensure the product water is well mixed to allow accurate measurements for the following steps.
2. Prepare the hemocytometer and the microscope to receive a sample of the product water
3. Dispense ~10-50 μL of product water into both counting chambers of the hemocytometer.
4. Use the microscope and attempt to count the number of observed yeast cells and microspheres. This may address **criteria 1 & 2.**
   1. If the yeast cells are hard to distinguish from the microspheres and test dust, follow steps 3 & 4 in the Challenge Water Makeup Procedures **without dilution.**
   2. If there are less than 5 cells in each square of the counting chamber, follow step 5-13.
5. Prepare the luer-lok filter by doing the following:
   1. Insert a 25 mm, 0.45 μm filter into the bottom half of the filter holder.  
      **Note: Use gloves and/or tweezers to handle filters to prevent contamination**  
      
   2. Place the silicone O-ring on top of the filter.  
      
   3. Screw the top piece of the filter holder.  
      
   4. Tighten down the filter holder with the use of two tongue and groove pliers.
6. Use a 100mL luer-lok syringe to collect 50 mL of the product water.
7. Attach the filter holder with filter onto the end of the filled syringe.
8. Push the product water through the syringe.
9. Push at least 100 mL of air through the filter to air in drying.
10. Remove the filter from the syringe.
11. Place the filter on a microscope slide.
12. Cover the filter with the counting grid.
13. Use the counting method listed [here](http://braukaiser.com/wiki/index.php/Microscope_use_in_brewing#counting).

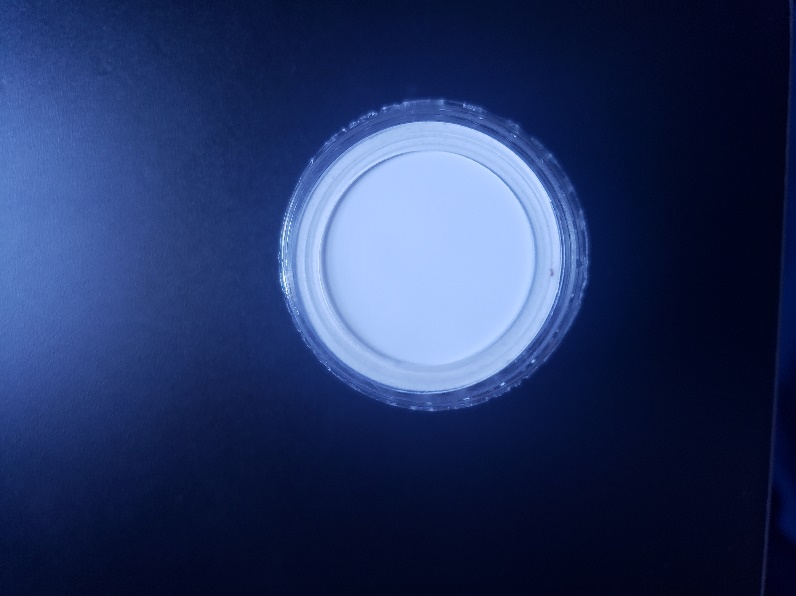
*Criterion #2: “Product meets or exceeds 3 log reduction of oocyst surrogate”*

**See criterion 1.**

**Category:** Product

**Method**: Cell count through hemocytometer or filter

**Procedure:**

1. Ensure the product water is well mixed to allow accurate measurements for the following steps.
2. Prepare the hemocytometer and the microscope to receive a sample of the product water.
3. Dispense ~10-50 μL of product water into both counting chambers of the hemocytometer.
4. Use the microscope and attempt to count the number of observed yeast cells and microspheres. This may address **criteria 1 & 2.**
   1. If the yeast cells are hard to distinguish from the microspheres and test dust, follow steps 3 & 4 in the Challenge Water Makeup Procedures **without dilution.**
   2. If there are less than 5 cells in each square of the counting chamber, follow step 5-13.
5. Prepare the luer-lok filter by doing the following:
   1. Insert a 25 mm, 0.45 μm filter into the bottom half of the filter holder.  
      **Note: Use gloves and/or tweezers to handle filters to prevent contamination**  
      
   2. Place the silicone O-ring on top of the filter.  
      
   3. Screw the top piece of the filter holder.  
      
   4. Tighten down the filter holder with the use of two tongue and groove pliers.
6. Use a 100mL luer-lok syringe to collect 50 mL of the product water.
7. Attach the filter holder with filter onto the end of the filled syringe.
8. Push the product water through the syringe.
9. Push at least 100 mL of air through the filter to air in drying.
10. Remove the filter from the syringe.
11. Place the filter on a microscope slide.
12. Cover the filter with the counting grid.
13. Use the counting method listed [here](http://braukaiser.com/wiki/index.php/Microscope_use_in_brewing#counting).

*Criterion #3: “Product meets turbidity specifications (≤ 5 NTU)”*

**Category:** Product

**Method:** The portable turbidity meter can measure the product’s turbidity.

**Procedure:**

1. A sample of the product water will be tested for turbidity to assess **criterion 3** using the Apera TN400 Turbidity meter. Please make sure the meter is calibrated and follow measurement procedures found on page 11 of the instruction manual.

*Criterion #4: “Product average flowrate meets or exceeds specifications*

*(≥ 18 mL/min)”*

**Category:** Cube Operation

**Method:** The amount of time it takes for the student’s cube to fill the required volume (90 mL) will determine the flowrate. Note that exceeding 40 mL/min will result in not meeting this requirement

**Procedure:**

1. The average flow rate for **criterion 4** will be calculated by diving the volume reached by the time it took to reach said volume.

*Criterion #5: “Product is within specified pH range (7 ± 0.5)”*

**Category:** Product

**Method:** Litmus paper will be used to estimate pH. If it is uncertain if the pH range is between 7 ± 1 or 7 ± 0.5, a pH probe will be used.

**Procedure:**

1. The pH will be determined via litmus paper test for **criterion 5**. If it is uncertain whether the pH is between 7 ± 1 or 7 ± 0.5, a pH probe will be used.

*Criterion #6: “Product chemical disinfectant concentration acceptable or non-chemical disinfection means provided for virus”*

**Category:** Product

**Method:** Students must incorporate a chemical disinfectant to be considered in this competition. For teams using chlorine disinfectant methods, the acceptable limit provided in the rubric will be used to judge a given team’s product water. Teams that use a non-chlorine based chemical disinfection must provide a testing method to measure the concentrations of their cube’s disinfectant, along with citations justifying the acceptable concentration limit for the non-chlorine based disinfection method.

**Procedure:**

1. A small sample of the product water is tested for chemical disinfectant concentration levels:
   1. For chlorine disinfectants, the safe drinking water upper limit according to the EPA should be detected: HOCl, OCl-, Cl2 (dissolved), and/or chloramines (total Cl): ≤4.0 mg/L; ClO2 or chlorine dioxide: ≤0.8 mg/L; Cl- or chloride: ≤250 mg/L.
      1. Take a clean cuvette and fill it with 10 mL of unreacted output water from a given team’s produced water. Place the plastic stopper and cap on the cuvette.
      2. Insert the cuvette into the chlorimeter and press **Zero**.
      3. Remove the cuvette.
      4. Add the content of 1 packet of total chlorine reagent to the filled cuvette. Shake gently for 20 seconds.
      5. Insert the cuvette into the chlorimeter and press **Read**. To skip the timer, press **Read** twice. Record the result in mg/L or ppm of chlorine.
   2. For hydrogen peroxide (H2O2) disinfectants, the safe drinking water upper limit according to the EPA should be detected: ≤50 ppm.
      1. Take the hydrogen peroxide test strip provided and dip it in a given team’s produced water sample.
      2. Record the detected hydrogen peroxide level based on the color of the strip using the color chart provided on the test strip container.

This will address **criterion 6**.

*Criterion #7: “Fills required volume (90 mL) of product before competitor in ‘The Duel’”*

**Category:** Cube Operation

**Method:** Takes less time than the competitor to fill the competition’s required volume (90 mL) of water.

**Procedure:**

1. During the duel, each cube’s average flow rate will be measured by independently (as opposed to using a timer for both cubes) timing the period of time in which the cube starts producing water to when the cube reaches the required volume. When the required volume is reached, the cube’s time will be stopped and recorded. The team with the lowest time in the duel receives points for **criterion 7**.

*Criterion #8: “Weighs less than its competitor in ‘The Duel’”*

**Category:** Cube Operation

**Method:** Weight will be determined in the safety pre-inspection. This is a comparison between the two competitors in a single duel,

**Procedure:**

1. During the safety inspection, each cube will be weighed-in by the safety judge.

*Criterion #9: “Consumes the least amount of electricity in ‘The Duel’”*

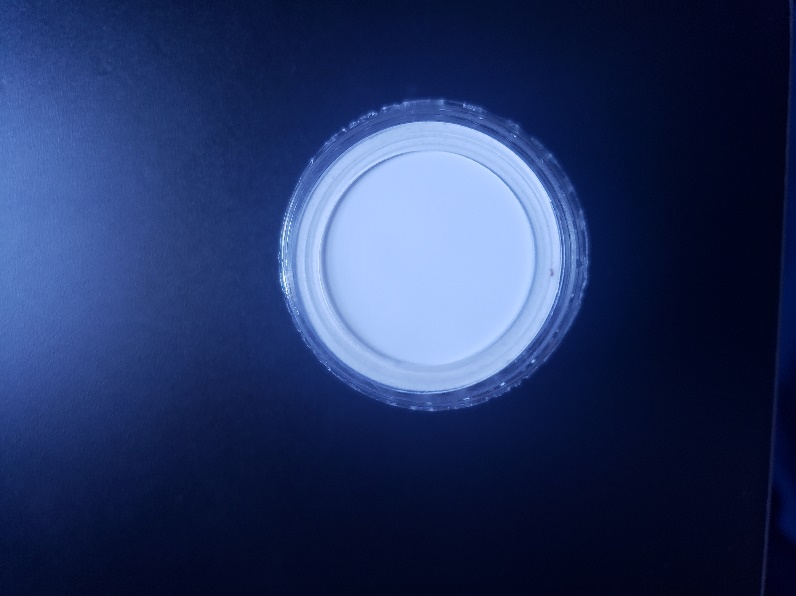
**Category:** Cube Operation

**Method:** Determined through an in-line power meter. Additionally, if a team is using a power plug (as opposed to banana plugs or post connectors), it must be a standard 120V US grounded electrical connector.

**Procedure**

1. Divide the power consumption by the time to determine the winner of **criterion 9** using the formula below

**Overall Testing Procedure**

1. During the duel, each cube’s average flow rate will be measured by independently (as opposed to using a timer for both cubes) timing the period of time in which the cube starts producing water to when the cube produces the required volume of 90 mL. When the required volume is reached, the cube’s time will be stopped and recorded. The team with the lowest time in the duel receives points for **criterion 7**.
2. The power consumption (in Watts) will be displayed on the in-line power meter. Record said value
3. Divide the power consumption by the time to determine the winner of **criterion 9** using the formula below
4. The average flow rate for **criterion 4** will be calculated by diving the volume reached by the time it took to reach said volume.
5. Each cube’s product will be collected and brought to the testing area for analysis.
6. The pH will be determined via litmus paper test for **criterion 5**. If it is uncertain whether the pH is between 7 ± 1 or 7 ± 0.5, a pH probe will be used.
7. A sample of the product water will be tested for turbidity to assess **criterion 3** using the Apera TN400 Turbidity meter. Please make sure the meter is calibrated and follow measurement procedures found on page 11 of the instruction manual.
8. A small sample of the water is taken to test for chemical disinfectant concentrations, if necessary. This will address **criterion 6**.
9. Ensure the product water is well mixed to allow accurate measurements for the following steps.
10. Prepare the hemocytometer and the microscope to receive a sample of the product water
11. Dispense ~10-50 μL of product water into both counting chambers of the hemocyotmeter.
12. Use the microscope and attempt to count the number of observed yeast cells and microspheres. This may address **criteria 1 & 2.**
    1. If the yeast cells are hard to distinguish from the microspheres and test dust, follow steps 3 & 4 in the Challenge Water Makeup Procedures **without dilution.**
    2. If there are less than 5 cells in each square of the counting chamber, follow step 13-27.
13. Prepare the luer-lok filter by doing the following:
    1. Insert a 25 mm, 0.45 μm filter into the bottom half of the filter holder.  
       **Note: Use gloves and/or tweezers to handle filters to prevent contamination**  
       
    2. Place the silicone O-ring on top of the filter.  
       
    3. Screw the top piece of the filter holder.  
       
    4. Tighten down the filter holder with the use of two tongue and groove pliers.
14. Use a 100mL luer-lok syringe to collect 50 mL of the product water.
15. Attach the filter holder with filter onto the end of the filled syringe.
16. Push the product water through the syringe.
17. Push at least 100 mL of air through the filter to air in drying.
18. Remove the filter from the syringe.
19. Place the filter on a microscope slide.
20. Cover the filter with the counting grid.
21. Adjust the slide holder in the microscope down, as shown in the image below:



1. Draw an X on a filter and use that filter to help focus the microscope.
2. Place the slide-filter sandwich on the microscope slide holder.
3. Use the grid on the microscope slide to count the number of yeast and microspheres (separately) per square.
4. Take an average count of 6 different squares for microspheres and yeast.
5. Multiply the average by ratio of the filter area and the observed area: 54.5
6. This number represents the total in the 50 mL, so simply divide this number by 50 to get the concentrations of yeast cells and microspheres, respectively. This will assess **criteria 1 & 2**.